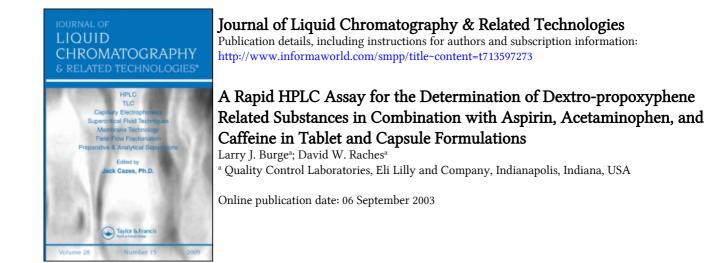
This article was downloaded by: On: 23 January 2011 Access details: Access Details: Free Access Publisher Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



**To cite this Article** Burge, Larry J. and Raches, David W.(2003) 'A Rapid HPLC Assay for the Determination of Dextropropoxyphene Related Substances in Combination with Aspirin, Acetaminophen, and Caffeine in Tablet and Capsule Formulations', Journal of Liquid Chromatography & Related Technologies, 26: 12, 1977 – 1990

To link to this Article: DOI: 10.1081/JLC-120021765 URL: http://dx.doi.org/10.1081/JLC-120021765

# PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.



JOURNAL OF LIQUID CHROMATOGRAPHY & RELATED TECHNOLOGIES<sup>®</sup> Vol. 26, No. 12, pp. 1977–1990, 2003

# A Rapid HPLC Assay for the Determination of Dextro-propoxyphene Related Substances in Combination with Aspirin, Acetaminophen, and Caffeine in Tablet and Capsule Formulations

Larry J. Burge\* and David W. Raches

Eli Lilly and Company, Quality Control Laboratories, Indianapolis, Indiana, USA

#### ABSTRACT

A 23 min gradient HPLC assay has been developed to simultaneously measure related substances of dextro-propoxyphene salts (napsalate, hydrochloride) and other actives that are routinely found in combination in capsules or tablets: aspirin, acetaminophen, and caffeine. This method has shown the ability to resolve all known active ingredients, impurities, and degradation products from the propoxyphene and other active drug parent peaks. All the propoxyphene related substances are quantified against a 1-point propoxyphene external standard prepared at 2% of the typical sample concentration. Propoxyphene related substance measurements occur at 210 nm. For acetaminophen combinations,

1977

DOI: 10.1081/JLC-120021765 Copyright © 2003 by Marcel Dekker, Inc. 1082-6076 (Print); 1520-572X (Online) www.dekker.com



<sup>\*</sup>Correspondence: Larry J. Burge, Eli Lilly and Company, Quality Control Laboratories, Drop 4923 Indianapolis, IN 46285, USA; E-mail: burge\_larry\_J@lilly.com.



wavelength switching is used starting at 280 nm to measure acetaminophen related substances, then switching to 210 nm for propoxyphene related substance measurements. Two sample solvents can be used for propoxyphene related substance quantitation, the second of which is used to provide enhanced room temperature stability-about one month-for aspirin containing formulations. This method provides a simple, one-step solution for related substance analysis rather than numerous separate assay methods for each active's related substances.

*Key Words:* HPLC; Propoxyphene; Acetaminophen; Aspirin; Caffeine; Salicyclic acid; Validation.

#### **INTRODUCTION**

Dextro-propoxyphene, as either hydrochloride or napsalate salt forms, is a widely prescribed analgesic alone, or in combination with other drugs: aspirin, caffeine, or acetaminophen. Official USP-NF analytical methods for the propoxyphene, alone or in combination, include labor-intensive sample extractions for GC analysis or various HPLC methods. The known related substances of propoxyphene-acetoxy analog and carbinol hydrochloride-utilize the labor intensive GC assay. Aspirin and caffeine combinations utilize two separate assay techniques: the labor-intensive GC assay for caffeine and a colorimetric assay for degraded aspirin to salicylic acid. Aspirin related, substances, 4-aminophenol and 4-chloroacetanailide, are determined by UV or TLC, respectively.<sup>[1]</sup>

A reverse phase HPLC method has been published for potency analysis of the dextropropoxyphene napsalate, caffeine, aspirin, and salicylic acid.<sup>[2]</sup> However, that method utilized samples prepared in methanol which is known to accelerate degradation of aspirin in solution.<sup>[3–14]</sup> Though that HPLC method claimed seven day stability of aspirin in methanolic solution, this is contrary to our experience and the experience of other Refs.<sup>[3–14]</sup> that indicate rapid deterioration to salicylic acid in less than 24 hours. No method has been found to simultaneously measure related substances of all the active drug components in a single method.

A 23 min reverse phase gradient HPLC method on C8 columns has been developed that provides quantitative related substance determination for propoxyphene alone or in combination with other drugs. Related substance determinations for the other active drugs can be determined concurrently. Two sample solvents are used: one for aspirin containing combinations that provide about one month room temperature stability for the aspirin, and a second for all other samples or combinations. Propoxyphene is stable in both sample



Marcel Dekker, Inc. 270 Madison Avenue, New York, New York 10016

#### **Determination of Dextro-proposyphene Related Substances**

Chemical names	Source		
Acetaminophen, aspirin, caffeine, dextro-propoxyphene salts, salicylic acid, and propoxyphene related substances (acetoxy analog and carbinol hydrochloride)	Corporate reference standards		
4-Aminophenol	Aldrich Chemical, Milwaukee, W		
4-Chloroacetanalide	Aldrich Chemical, Milwaukee, W		
Hydroquinone	Aldrich Chemical, Milwaukee, W		
Acetylsalicylsalicylic acid	Acros Organics, NJ		
Salicylsalicylic acid	Acros Organics, NJ		
Hydrochloric acid, 0.02 N (prepared from conc. reagent)	Fisher, FairLawn, NJ		
Potassium phosphate monobasic, HPLC grade	Fisher, FairLawn, NJ		
Water, HPLC grade	Fisher, FairLawn, NJ		
Acetonitrile, HPLC grade	Fisher, FairLawn, NJ		
O-Phosphoric acid, HPLC grade	Fisher, FairLawn, NJ		
Formic acid, GR	EM Science, Gibbstown, NJ		
Citric acid monohydrate	Mallinckrodt, Paris, KY		

Table 1. Reagents and chemicals.

solvents for at least seven days at room temperature; however, acetaminophen solutions must be stored at  $5^{\circ}$ C to achieve four day stability per ICH Guidelines on stability.<sup>[15]</sup>

#### **EXPERIMENTAL**

# **Reagents and Chemicals (Table 1)**

Sample Solution Preparation

Primary sample solution: 60% hydrochloric acid (0.02 N)/40% acetonitrile. Alternate sample solution (for aspirin stability): 99 parts acetonitrile, 1 part formic acid, and 0.1 part citric acid.

# Mobile Phase Preparation

Mobile phase A—a 50 mM solution of potassium phosphate monobasic was prepared in purified water. The solution was adjusted to pH  $2.4 \pm 0.1$ 



Copyright © 2003 by Marcel Dekker, Inc. All rights reserved.



using phosphoric acid. Mobile phase B-60% acetonitrile/40% mobile phase A.

# Instrumentation and Chromatographic Conditions

A HPLC system comprised of a Waters 2690 Separations Module (Alliance System), Waters 2487 variable UV detector, a PE Nelson box interface, and a Hewlett Packard HP1000 data system was used for all experiments. Two alternate Agilent HPLC columns were used, 5  $\mu$ m particle, 4.6  $\times$  150 mm; a Zorbax SB C8 or an Eclipse XDB C8. The following gradient program was applied (Table 2).

Initially, all analysis was done at 210 nm to maximize sensitivity. An alternate wavelength switching program was adopted for analysis of acetaminophen and its related substances starting at 280 nm, then switching back to 210 nm for analysis of other active drugs related substances.

Since analysis is being done at 210 nm for most of the analytes, extreme care must be taken to ensure that glassware is clean and reagents are HPLC grade or better. Any contamination can affect the baseline quality, especially later in the gradient where we wish to quantify very small propoxyphene related substances. Routinely during our development studies, we incurred significant baseline shifts due to glassware. The corrective actions were to remove mobile phases, methanol wash and thoroughly dry the mobile phase flasks, methanol wash the HPLC column to remove all contaminants, then remake fresh mobile phases and check baselines. This whole process is the hardest part of this low UV assay, but controllable with care.

Table 2.	Gradient conditions:	linear
gradient i	from A to B.	

Time	Flow	%A	%B	
0.0	1.0	85	15	
12.0	1.0	25	75	
13.0	1.0	0	100	
18.0	1.0	0	100	
18.5	2.0	85	15	
22.0	2.0	85	15	
22.1	1.0	85	15	
23.0	1.0	85	15	



Copyright @ 2003 by Marcel Dekker, Inc. All rights reserved

270 Madison Avenue, New York, New York 10016

#### **Determination of Dextro-propoxyphene Related Substances**

#### **Preparation of Standard Solutions**

Separate propoxyphene standards are prepared for each salt form being analyzed. Approximately 0.5 mg/mL solutions as the propoxyphene base were prepared for each salt form (for HCl salt, MW = 375.98, prepare about 50–55 mg propoxyphene hydrochloride per 100 mL in primary sample diluent; for napsalate salt, MW = 564.48, prepare about 90–100 mg propoxyphene napsalate per 100 mL). A 1/50 dilution is made of each propoxyphene stock solution in the same sample solvent to achieve 2% standards for quantitation of propoxyphene related substances.

Separate stock standards for acetaminophen and caffeine were prepared in primary sample solvent and diluted to achieve about 1% standards for quantitation of their related substances. Separate 1 mg/mL aspirin and salicylic acid stock standards are prepared in the alternate sample solvent and diluted 1/100 to quantify aspirin related substances. (Note: propoxyphene standards can also be prepared in the alternate sample solvent, but caffeine and acetaminophen chromatography peak distortion prevents use of this solution use for those standards).

#### **Preparation of Standard Curves**

Separate calibration curves were made for each active drug and their known related substances to determine response factors of all components relative to propoxyphene, and to determine the limit of quantitation (LOQ)/limit of detection (LOD) of propoxyphene. A propoxyphene hydrochloride standard solution in primary sample solvent was diluted across a concentration range of 0.8–40 mcg/mL (0.16–8% typical sample concentrations of propoxyphene and bracketing the routine 2% standard used for 1-point quantitation). Similar ranges of other active drugs and their individual related substances (except aspirin and salicylic) were prepared in primary sample solvent. For aspirin and salicylic comparisons to propoxyphene, all stock solutions including propoxyphene and standard curves were made in alternate sample solvent.

# Preparation of Capsule and Tablet Formulations for Analysis

Four different formulations manufactured by Eli Lilly and Co. were prepared in sample solvents (aspirin containing tablet prepared in both solvents using the alternate sample solvent to quantify both aspirin and propoxyphene related substances, then the primary sample solvent to again quantify propoxyphene related substances along with caffeine). Related



Active drug	Ingredient levels (mg)				
	Capsule #1	Capsule #2	Tablet #1	Tablet #2	
Propoxyphene	65.00	65.65	101.00	50.50	
Acetaminophen	None	None	None	325.00	
Caffeine	None	32.72	None	None	
Aspirin	None	389.00	None	None	

Table 3. Formulations tested.

substance quantitation of propoxyphene in both solvents was compared within method precision studies to ensure no statistical difference in results occurred. The ingredient compositions given in Table 3 were tested.

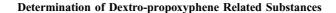
## **Preparation of Placebo Samples**

Excipient ingredients in each tablet or capsule formulation were prepared, without the active drugs, in sample solvents to simulate levels expected in the 0.5 mg/mL propoxyphene base equivalent sample preparations. Each placebo was profiled against its respective active drug sample preparation to assess interferences. Placebo mixtures were also exposed to heat stressing (80°C for three and seven days) to determine if any impurities due to the placebos could be generated that would interfere with analysis of active drug related substances. Additionally, each color ingredient and other tablet coating ingredients were evaluated for interferences.

# **RESULTS AND DISCUSSION**

Our United Kingdom affiliate had established a potency method for acetaminophen and propoxyphene combined in one tablet formulation that used wavelength switching from 280 nm for acetaminophen analysis to 205 nm for propoxyphene. That method utilized a  $4.6 \text{ mm} \times 25 \text{ cm}$  Kromasil C18 with 40% acetonitrile in a 12 millimolar, pH 2.5 potassium phosphate buffer at 1 mL/min isocratic analysis. Though propoxyphene related substances (acetoxy and carbinol compounds) and acetaminophen related substances (4-aminophenol and 4-chloroacetanailde) are resolved from the active drugs in this method, separate methods are used to analyze those compounds. The acetoxy and carbinol are analyzed using a perchloric acid buffer at pH 2 on a C18 column and 217 nm UV detection, while 4-aminophenol is analyzed

Marcel Dekker, Inc. 270 Madison Avenue, New York, New York 10016



*Table 4.* Typical retention times of active ingredients, related substances, impurities, and degradation products in propoxyphene formulations.

Peak identity	Typical retention time (sec)	
Void time	60	
Capsule #2 placebo peak	85	
4-Aminophenol, degradation of acetaminophen, starting material in synthesis	95	
Formic acid, sample diluent component	112	
Acetic acid, aspirin degradation	119	
Hydroquinone-solution degradation peak of aminophenol and acetaminophen	202	
Acetaminophen	252	
Caffeine	347	
Orange color mixture for tablet coating	376	
Napsalate peak of propoxyphene salt	419	
Aspirin	526	
Salicylic acid	578	
Carbinol derivative of propoxyphene	668	
4-Chloroacetanalide, synthesis int. of acetaminophen	690	
Acetoxy derivative of propoxyphene	711	
Propoxyphene	775	
Acetylsalicylsalicylic acid, aspirin deg. Late peak #1	849	
Salicylsalicylic acid, aspirin deg. Late peak #2 Late aspirin deg. Peaks 3–8	905 949–1150	

using butane sulfonate ion-pairing with a C18 column and 272 nm UV detection. The 4-chloroacetanalide in acetaminophen is tested using a thin layer chromatography method.

We were interested in developing a single method to be used for the determination of related substances in each of the four propoxyphene formulations. The conditions above, for propoxyphene and acetaminophen potency gave poor chromatography of aspirin and salicylic, insufficient retention of the 4-aminophenol, and incomplete elution of some of the aspirin degradation analogs. Shorter carbon chain reverse phase columns and gradient optimization was pursued to get the fastest run time possible, while providing good chromatography of all the active compounds and their related substances. Both methanol and acetonitrile in pH 2.5 potassium phosphate gradients were evaluated on various C8 columns at 210 nm, a wavelength where we appeared to have similar response of propoxyphene and its related substances, while providing good sensitivity for other target compounds.

Compound	Response factor vs. propoxyphene		
Response at 210 nm			
Acetoxy	1.1		
Carbinol	1.1		
Acetaminophen	1.1		
Caffeine	2.5		
Aspirin	0.9		
Salicylic acid	3.6		
4-Aminophenol	0.9		
4-Chloroacetanalide	1.0		
Response at 280 nm			
Acetaminophen	0.27		
4-Aminophenol	0.20		
Caffeine	0.66 <sup>a</sup>		

Table 5. Relative response factors.

<sup>a</sup>Response is  $2.45 \times \text{acetaminophen}$ , similar to 210 nm.

Ultimately, a  $10 \,\mu\text{L}$  sample injection onto 15 cm Zorbax SB C8 and XDB C8 columns with an acetonitrile/phosphate buffer gradient was chosen for fastest and best chromatography of target compounds.

# Selectivity

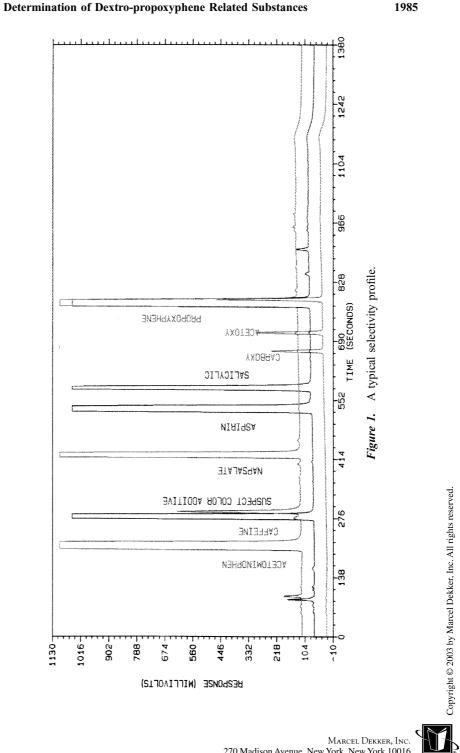
Table 4 summarizes typical retention times of active drugs, known related substances, degradation products, excipients, and tablet coatings.

The chromatograms given in Figs. 1 and 2 profile routine formulations and some heat stressed samples to force impurities for selectivity confirmation. These analyses are all done at 210 nm, only, without using wavelength switching for acetaminophen related substances.

Aspirin standards or a formulation containing aspirin was heat degraded at 80°C for three and seven days to promote the generation of salicylic acid and other aspirin degradation peaks in Fig. 2. This heat degraded sample is used as part of system suitability to confirm resolution of aspirin analogs from the propoxyphene peak and to assess resolution of the aspirin–salicylic acid critical pair.

Solution pH studies show that the critical pair resolution is best below a pH of 3 in typical reverse phase systems.<sup>[16]</sup> We evaluated the impact of low

Marcel Dekker, Inc. 270 Madison Avenue, New York, New York 10016

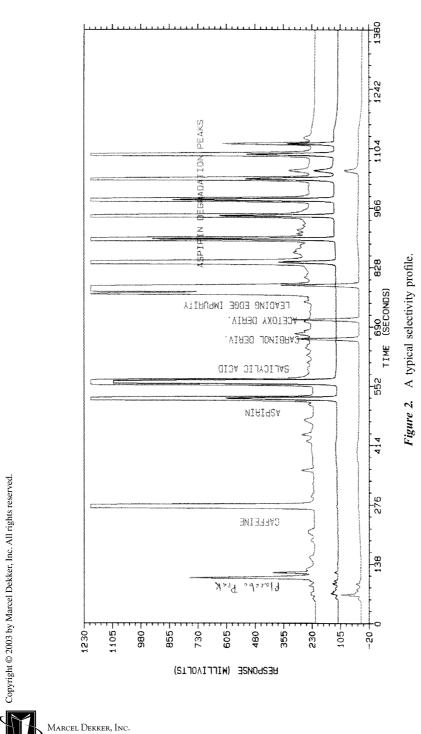


ੂ ORDE

REPRINT

Marcel Dekker, Inc. 270 Madison Avenue, New York, New York 10016

Downloaded At: 20:00 23 January 2011





1986



Marcel Dekker, Inc. 270 Madison Avenue, New York, New York 10016

#### **Determination of Dextro-propoxyphene Related Substances**

1987

pH range (about 2–3) on all actives, impurities, degradation products, and excipient components. Only the aspirin–salicylic acid pair was affected with improved resolution as pH was lowered. We locked in on a pH of  $2.4 \pm 0.1$  for routine analysis to maximize resolution without broaching the pH limit of the analytical columns at pH 2.

## Linearity

Response factors of the active drugs and their known related substances were determined at 210 nm relative to proposyphene using a ratio of curve slopes against the proposyphene curve. Also, response factors for acetaminophen, aminophenol, and caffeine were determined at 280 nm (Table 5).

The LOD and LOQ were determined for propoxyphene hydrochloride using two separate techniques:  $3 \times \text{noise}$  estimate from typical chromatogram = LOD and  $10 \times \text{noise} = \text{LOQ}$ ; alternatively, LOQ determined from the standard curve =  $10 \times \text{STD}$  deviation of curve/mean response. Both estimates of LOQ provided similar results of 0.01% for sample preparations of 0.5 mg/mL, well below ICH Guidelines recommendation of 0.1% LOQ based on daily dose of drug.

For the typical 0.5 mg/mL proposyphene sample preparations, the LOQ of free salicylic in the formulations is about 20 fold more sensitive due to the approximate 7:1 ratio of aspirin to proposyphene in capsules tested and the 3+ response factor for salicylic acid. This method is very sensitive for free salicylic determinations.

## Stability

Sample solution stability of all the active drugs and their known related substances were checked in primary sample solvent at room temperature. A change of less than 0.1–0.2% total related substances in the active drugs was accepted as an indicator of solution stability. All four propoxyphene formulations, propoxyphene and caffeine standard solutions were stable beyond seven days at room temperature.

Aspirin is not stable in the aqueous solvent, but in the alternate sample solvent aspirin sample solutions were stable at room temperature for 3–4 weeks. Salicylic acid begins forming almost immediately when aspirin is dissolved in primary sample solvent.

Acetaminophen in primary sample solvent degrades to aminophenol, which then degrades to hydroquinone at room temperature. However, if samples are stored refrigerated at 5°C, stability is achievable for up to four days. The hydroquinone degradant was determined by GC/MS in aged aminophenol solutions and confirmed with spiking experiments. This



degradation pathway appears to be unique. An organic text investigated for aminophenol in the presence of HCl and acetonitrile only suggested the Pinner synthesis (alcoholysis of nitriles using conc. HCl) to produce carbamides.<sup>[17]</sup> We only use 0.02 N HCl solution with acetonitrile in the primary sample solution and room temperature aging.

Caffeine generated no related substances when stored for weeks in solution. Additionally, when caffeine was heat stressed at 80°C for three and seven days and put in solution, no related substances were detected.

#### Accuracy

Placebo samples of each formulation were prepared and wet spiked with standard solutions to assess recovery. At five times typical placebo ingredient ratio to propoxyphene levels, 99-101% recovery was achieved except with the formulation containing a clay product-Kaolin. Quantitative recovery (>95%) was not achievable until Kaolin to propoxyphene ratio neared a 2:1 typical mix. One hundred percent was recovered in the typical formulation mix.

# Precision

Both intermediate precision and repeatability experiments were done using all formulations. Additionally, precision of propoxyphene was checked using the alternate sample diluent to compare to primary sample diluent for capsule #2 containing aspirin (Table 6).

# CONCLUSION

This simple, 23 min HPLC method provides a quick, quantitative determination of propoxyphene related substances in solid dose formulations with stability and limits of quantitation that exceed ICH Guidelines. Additionally, this method can be used to determine related substances of other active drugs formulated in combination with the propoxyphene concurrently with the

Table 6. Precision estimates of propoxyphene related substance measurements.

	Capsule #1- diluent 1	Capsule #2- diluent 1	Capsule #2- diluent 2	Tablet #1	Tablet #2
Repeatability	3.13%	5.50%	1.66%	3.78%	1.71%
Int. precision	3.60%	5.67%	4.43%	2.75%	5.11%

Marcel Dekker, Inc. 270 Madison Avenue, New York, New York 10016

#### **Determination of Dextro-propoxyphene Related Substances**

1989

propoxyphene. Enhanced stability of aspirin, up to one month at room temperature, allows for accurate quantitation of free salicylic acid formed over time in aged formulations.

#### ACKNOWLEDGMENTS

We thank Patrick Jansen, Mathew Clemens, and Wayne Taylor for their LC/MS and GC/MS analysis supporting identification of impurities; Don Risley for his scientific review and inputs; and Simon Mills and Alan Reeves of our United Kingdom laboratories for their assistance in early phases of this method development.

# REFERENCES

- 1. United States Pharmacopeia 25-National Formulary 20; United States Pharmacopeial Convention, Inc.: Rockville, MD, 2001; 16–21, 274–275, 1466–1474.
- Jalal, I.M.; Sa'sa', S.I. Simultaneous determination of dextropropoxyphene napsalate, caffeine, aspirin, and salicylic acid in pharmaceutical preparations by reversed-phase HPLC. Talanta **1984**, *31* (11), 1015–1017.
- Montgomery, E.R.; Taylor, S.; Segretario, J.; Engler, E.; Sebastian, D. Development and validation of a reverse-phase liquid chromatographic method for analysis of aspirin and warfarin in a combination tablet formulation. J. Pharm. Biomed. Anal. **1996**, *15*, 73–82.
- Bevitt, R.N.; Mather, J.R.; Sharman, D.C. Minimisation of salicylic acid formation during preparation of aspirin products for analysis by highperformance liquid chromatography. Analyst **1984**, *109*, 1327–1329.
- Xu, X.; Stewart, J.T. HPLC methods for aspirin-caffeine-butalbital and acetaminophen-caffeine-butalbital mixtures in tablet dosage forms using non-porous octadecylsilane columns. J. Liq. Chrom. & Rel. Technol. 2000, 23 (5), 769–779.
- 6. Luzzi, L.A.; Whitworth, C.W.; Jun, H.W. Solubilized Aspirin. US Patent 3842170, April 13, 1973.
- Galante, R.N.; Visalli, A.J.; Grim, W.M. Stabilized normal-phase highperformance liquid chromatographic analysis of aspirin and salicylic acid in solid dosage forms. J. Pharm. Sci. 1984, 73 (2), 195–197.
- Climent Morato, M.D.; Rico Selas, M.I.; Jiminez Gomez, S. Stability of acetylsalicylic acid in alcoholic solutions as a preliminary phase for gel chromatographic analysis. An. R. Acad. Farm. 1978, 44 (3), 363–370.

# 

#### **Burge and Raches**

- 9. Choudhury, S.; Mitra, A.K. Kinetics of aspirin hydrolysis and stabilization in the presence of  $\beta$ -cyclodextran. Phar. Res. **1993**, *10* (1), 156–159.
- Chavkin, L.; Mackles, L. Shelf-stable Aspirin Solutions for Topical Application. US Patent 4975269, December 4, 1990.
- Carstensen, J.T.; Attarchi, F. Decomposition of aspirin in the solid state in the presence of limited amounts of moisture. J. Pharm. Sci. 1988, 77 (4), 314–317.
- Gharbo, S.A.; Williamson M.J. Degradation of aspirin in solutions. Comments Drug. Dev. Ind. Pharm. 1986, 12 (6), 927–929.
- Burghart, W.; Burghart, K. Method for Producing Stable acetylsalicylic Acid Solutions. Austrian Patent WO 9903474, January 28, 1999.
- Kornblum, S.S.; Zoglio, M.A. Pharmaceutical heterogeneous systems. I. Hydrolysis of aspirin in combination with tablet lubricants. J. Pharm. Sci. 1967, 56 (12), 1569–1575.
- 15. ICH Steering Committee, ICH Harmonized Tripartite Guideline: Impurities In New Drug Products. *International Conference on Harmonization of Technical Requirements for Registration of Pharmaceuticals for Human Use*, recommended for adoption at Step 4 of the ICH process, November 6, 1996.
- Menouer, M.; Bouabdellah, F.; Ghernati, H.M.; Guermouche, M.H. HPLC of acetylsalicylic (ASA) and salicylic acids (SA): optimal conditions for trace analysis of SA and ASA. J. HRC CC 1982, 5, 267–268.
- 17. March, J. Advanced Organic Chemistry: Reactions, Mechanisms, and Structure, 4th Ed.; John Wiley and Sons, Inc.: 1992; 892 pp.

Received December 1, 2002 Accepted December 31, 2002 Manuscript 6037

1990

Marcel Dekker, Inc. 270 Madison Avenue, New York, New York 10016